

# Quantitative In Vivo Redox Sensors Uncover Oxidative Stress as an Early Event in Life

Daniela Knoefler,<sup>1,5</sup> Maïke Thamsen,<sup>1,5</sup> Martin Koniczek,<sup>2,4</sup> Nicholas J. Niemuth,<sup>3</sup> Ann-Kristin Diederich,<sup>1</sup> and Ursula Jakob<sup>1,\*</sup>

<sup>1</sup>Department of Molecular, Cellular, and Developmental Biology

<sup>2</sup>Department of Radiation Oncology

<sup>3</sup>Cellular and Molecular Biology Training Program  
University of Michigan, Ann Arbor, MI 48109, USA

<sup>4</sup>Experimental Radiation Oncology, Medical Center Mannheim-Heidelberg University, 68167 Mannheim, Germany

<sup>5</sup>These authors contributed equally to this work

\*Correspondence: [ujakob@umich.edu](mailto:ujakob@umich.edu)

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## SUMMARY

Obstacles in elucidating the role of oxidative stress in aging include difficulties in (1) tracking in vivo oxidants, in (2) identifying affected proteins, and in (3) correlating changes in oxidant levels with life span. Here, we used quantitative redox proteomics to determine the onset and the cellular targets of oxidative stress during *Caenorhabditis elegans*' life span. In parallel, we used genetically encoded sensor proteins to determine peroxide levels in live animals in real time. We discovered that *C. elegans* encounters significant levels of oxidants as early as during larval development. Oxidant levels drop rapidly as animals mature, and reducing conditions prevail throughout the reproductive age, after which age-accompanied protein oxidation sets in. Long-lived *daf-2* mutants transition faster to reducing conditions, whereas short-lived *daf-16* mutants retain higher oxidant levels throughout their mature life. These results suggest that animals with improved capacity to recover from early oxidative stress have significant advantages later in life.

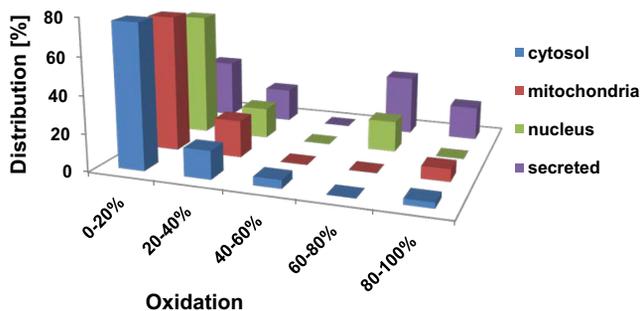
## INTRODUCTION

The oxidative stress theory of aging postulates that oxidative damage to cellular macromolecules, caused by the progressive accumulation of reactive oxygen species (ROS), contributes and possibly even leads to the decline in physiological functions observed in aging organisms (Finkel and Holbrook, 2000; Harman, 1956). Since its inception, extensive correlative evidence has been collected that corroborates this popular theory. For instance, it has been shown that oxidative damage to proteins, lipids, and DNA increases with age (Finkel and Holbrook, 2000) and that interventions that delay aging (e.g., reduced caloric intake) decrease the extent of oxidative damage and mediate the rapid removal of damaged macromolecules through proteol-

ysis and autophagy (Cavallini et al., 2008). Moreover, one of the unifying features that distinguishes many long-lived mutant invertebrates and rodents from their wild-type cohorts appears to be a significant increase in oxidative stress resistance (Salmon et al., 2005). However, the validity of the free radical theory of aging recently came under dispute when a series of genetic studies, particularly those in mice, failed to generate conclusive results about the role of ROS in aging (Perez et al., 2009). These findings suggested a more complex aging mechanism, potentially driven by more subtle changes in ROS levels or the cellular redox state.

One aspect of redox biology that has long been overlooked when testing the free radical theory of aging is the fact that ROS, like peroxide, also play important regulatory roles as intracellular signaling molecules. While high levels of peroxide are thought to be toxic, low levels of peroxide, which are continuously produced during mitochondrial respiration and other processes, set the pace of numerous metabolic and signaling pathways in the cell (D'Autreaux and Toledano, 2007). Proteins that are regulated by peroxide and potentially other reactive oxygen and nitrogen species commonly contain highly oxidation-sensitive cysteine residues whose thiol oxidation status controls the protein's activity and, by extension, the pathway that the protein is part of (Cross and Templeton, 2006). Given this fact, it is thus very likely that depletion of pro-oxidants by either dietary antioxidants or genetic manipulation profoundly impacts development, differentiation, and stress responses and, when applied at the wrong stage in life, potentially outweighs the beneficial effects.

We decided to take a different approach in evaluating the role that ROS play in life span. We used recently developed quantitative probes to determine when ROS accumulate, which proteins and pathways they affect, and whether a correlation exists between the onset and extent of endogenous oxidative stress and the life span of the organism. We chose *C. elegans* as our model system because it is a well-established eukaryotic aging model and is well-suited for redox proteomic analyses and the application of fluorescent biosensors (Johnson, 2008; Kumsta et al., 2010). To monitor global changes in the cellular redox environment, we applied the quantitative redox proteomic technique OxICAT at distinct time points in *C. elegans* life span to



**Figure 1. In Vivo Thiol Oxidation Status of *C. elegans* Proteins**

Wild-type N2 worms were synchronized and cultivated at 15°C. At day 2 of adulthood, worms were lysed, and the protein thiol oxidation status was determined using the differential thiol trapping technique OxICAT. Proteins are categorized by their oxidation status and subcellular localization, according to WormBase (<http://www.wormbase.org/>). The complete list of proteins and their respective oxidation states with standard deviations can be found in Table S1.

determine the thiol redox status of almost 140 different proteins. In parallel, we employed the chromosomally encoded hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) sensor protein HyPer (Belousov et al., 2006) as a tissue-specific readout for changes in endogenous peroxide levels. By using these quantitative redox sensors, we discovered that *C. elegans* is exposed to high oxidant levels at two distinct stages in life: during development and during aging. Comparative analysis with long- and short-lived mutants of the insulin/IGF-1 signaling (ILS) pathway revealed that long-lived mutant worms recovered faster from increased developmental ROS levels and reached lower steady-state redox levels during their reproductive period than short-lived worms, which failed to completely restore redox homeostasis. These results suggest that changes in the cellular redox homeostasis, encountered at a very early stage in life, determine subsequent redox levels and potentially the life span of organisms.

## RESULTS

### The *C. elegans* Redoxome: Establishing the Redox Baseline

We recently developed a redox proteomic technique, termed OxICAT, which allows us, in a single experiment, to determine the in vivo oxidation status of hundreds of different protein thiols, many of which are redox sensitive (Kumsta et al., 2010; Leichert et al., 2008). We reasoned that by monitoring the in vivo redox state of protein thiols over the life span of *C. elegans*, we should obtain information about temporal and spatial changes in cellular redox homeostasis and identify processes and pathways that might be affected by the prevailing redox conditions. With OxICAT, we use the quantitative properties of the thiol-reactive isotope-coded affinity tag (ICAT) to differentially label in vivo reduced and in vivo oxidized protein thiols (see Figure S1 available online). HPLC is used to separate the ICAT-labeled peptides, followed by mass spectrometry (MS) and tandem MS/MS, which allow us to identify the thiol-containing peptides and quantify their in vivo oxidation status (Leichert et al., 2008).

Due to its ratio-based nature, OxICAT is independent of the protein amount present in cells and tissues, making it ideally suited to monitor changes in protein oxidation over the life span of the organism.

We applied OxICAT to analyze the redox status of proteins in synchronized wild-type *C. elegans* on day 2 of adulthood. This analysis in young adults was intended to serve as a reference point for all subsequent studies so that relative changes in the redox status of the proteins could be determined at different time points during *C. elegans* life span. As shown in Table S1, we reproducibly determined the redox status of 170 thiol-containing peptides, representing 137 different *C. elegans* proteins. Database analysis revealed that we identified the redox status of numerous ubiquitously expressed proteins (e.g., HSP-1, 17 ribosomal proteins) as well as proteins selectively expressed in body wall muscle (e.g., ANC-1, DIM-1, myosin-3), intestine (e.g., vitellogenin-6, neprilysin), nervous system (e.g., degenerin), and pharynx (e.g., myosin-2, myosin-4, annexin). Of the identified protein thiols, 65% exhibited oxidation states of less than 20%, with most of them being less than 10% oxidized at this point in life. These protein thiols were found predominantly in cytosolic and mitochondrial proteins (Figure 1). About 25% of our identified peptides contained cysteines with oxidation levels between 20% and 60% (Figure 1). This partially oxidized group of proteins included enzymes involved in glucose metabolism (e.g., alcohol dehydrogenase, fructose-bisphosphate aldolase) and ATP-homeostasis (e.g., nucleoside diphosphate kinase, vacuolar ATP synthases), as well as proteins involved in motility (e.g., UNC-87, paramyosin), signal transduction (e.g., phosphoinositide kinase AGE-1), and protein homeostasis (e.g., ATP-dependent chaperone CDC-48) (Table S1). Many of these proteins have been previously shown to contain redox-sensitive cysteines (Kumsta et al., 2010) (Table 1), suggesting that they are partially oxidized in response to either local or global ROS accumulation or due to changes in cellular redox homeostasis. Less than 10% of our identified protein thiols were oxidized greater than 80%. Most of these highly oxidized thiols belonged to secreted proteins with known disulfides (Figure 1).

### Monitoring Changes in Protein Thiol Redox State during the Life Span of *C. elegans*

To investigate the redox state of the identified thiol-containing proteins over the life span of *C. elegans*, we took aliquots of ~100,000 worms from a synchronized wild-type population during early (i.e., L2) and late (i.e., L4) development and at days 2, 8, and 15 of adulthood for subsequent OxICAT analysis. Each of the time points was selected based on the fact that they represent distinct physiological stages in *C. elegans*. The larval stages L2 and L4 represent early and late postembryonic developmental stages of *C. elegans*. Worms transition to their reproductive phase, which peaks around day 2–3 of adulthood. At day 8, most animals have ceased reproduction, yet worms still reveal juvenile mobility and pharyngeal pumping. By day 15, significant mobility defects are observed in the majority of worms; however, the mean survival of the population is still about 80%. We separated live worms from eggs and any dead worms by sucrose flotation, lysed the worms in trichloroacetic acid to maintain the in vivo redox status of the protein thiols, and

## Molecular Cell

### Oxidative Stress, an Early Event in Life

**Table 1. Oxidation Status of Select Proteins during *C. elegans* Life Span**

Protein (Affected Cysteine)	Accession Number	Tissue <sup>g</sup>	Average Oxidation Status <sup>a</sup>			Oxidation Sensitive
			Dev <sup>b</sup>	Young <sup>e</sup>	Old <sup>f</sup>	
Tubulin (alpha-2 chain) (C314)	CE09692	N	23 ± 6	7 ± 2	ND	Yes <sup>i</sup>
GTP-binding protein (TAG-210) (C54)	CE14708	I	20 ± 1 <sup>c</sup>	7 ± 6	ND	
Protein CE32871 (C374)	CE32871		26 ± 3	12 ± 3	ND	
Proteasome alpha subunit (PAS-3) (C74)	CE30307		26 ± 6 <sup>c</sup>	13 ± 1	ND	
Heat shock protein 70 HSP-1 (C243)	CE09682	P, I, BW	31 ± 2 <sup>c</sup>	4 ± 2	9 ± 4	Yes <sup>i</sup>
Aspartyl-tRNA synthetase (C233)	CE00015		27 ± 8	6 ± 2	17 ± 11	
Vacuolar ATP synthase (subunit A) (C218)	CE22210	I, BW	28 ± 5	10 ± 2	11 ± 4	Yes <sup>h</sup>
T-complex protein 1 (subunit zeta) (C517)	CE01234	U	30 ± 4	12 ± 2	26 ± 14	Yes <sup>i</sup>
40S ribosomal protein S28 (C22)	CE21842	U	20 ± 3	6 ± 3	29 ± 4	Yes <sup>h</sup>
DIM1 (C234)	CE09308	BW	18 ± 5	8 ± 3	33 ± 14	Yes <sup>h</sup>
COPII coatomer (subunit SAR-1) (C174)	CE07622		23 ± 5	10 ± 2	25 ± 7	
Methylcrotonyl-CoA carboxylase (C212)	CE00136	P	26 ± 5	10 ± 6	22 ± 2	
40S ribosomal protein S12 (C114)	CE26896	U	30 ± 5	13 ± 4	34 ± 6	Yes <sup>h</sup>
Nepriylsin (C41)	CE43217	I	66 ± 4 <sup>c</sup>	22 ± 6	62 ± 6	
Serine hydroxymethyltransferase MEL-32 (C391)	CE01130	I, H	63 ± 3 <sup>d</sup>	31 ± 4	44 ± 3	Yes <sup>h</sup>
UNC-87 (C472)	CE36924	BW, M	68 ± 6	40 ± 2	54 ± 6	
Vitellogenin-6 (C468)	CE28594	I	89 ± 5	65 ± 4	80 ± 8	
Chitinase (C738/747)	CE32592	G	94 ± 5	68 ± 1	91 ± 2	Yes <sup>i</sup>
Vitellogenin-3 (C626)	CE20900	I	ND	10 ± 2	76 ± 9	
RNA helicase CGH-1 (C336)	CE00839	G	ND	22 ± 4	45 ± 10	
60S ribosomal protein L22 (C27)	CE04102	U	8 ± 2	6 ± 2	19 ± 5	Yes <sup>h</sup>
Protein K07C5.4 (C390)	CE06114		12 ± 4 <sup>c</sup>	9 ± 5	28 ± 8	
Small molecule methylase (C28H8.7) (C224)	CE01829		2 ± 2	12 ± 1	26 ± 6	
Actin-2 (C258)	CE13150	BW, N	16 ± 10 <sup>c</sup>	16 ± 3	30 ± 4	Yes <sup>i</sup>
Glutamyl-tRNA synthetase (C377)	CE06580		14 ± 5 <sup>c</sup>	22 ± 2	33 ± 3	
Translation initiation factor 5A (C111)	CE37787	G	17 ± 3 <sup>c</sup>	27 ± 6	47 ± 2	Yes <sup>h</sup>
Nucleoside diphosphate kinase NDPK (C109)	CE09650	U	35 ± 6	28 ± 4	47 ± 8	Yes <sup>h</sup>
Nucleoside diphosphate kinase NDPK (C117)	CE09650	U	36 ± 2	30 ± 5	54 ± 13	Yes <sup>h</sup>
Phosphoinositide 3-kinase AGE-1 (C710)	CE43431	I, N	56 ± 4	50 ± 12	72 ± 5	

ND, peptide not reproducibly identified at this stage.

<sup>a</sup>Average oxidation of at least three individual replicates.

<sup>b</sup>Thiol oxidation status of peptides in L4 stage.

<sup>c</sup>Thiol oxidation status of peptide in L2 stage was used if corresponding peptide was not identified in L4 larva.

<sup>d</sup>Thiol oxidation status of peptide in L2 stage was also used if thiol oxidation in L2 was substantially higher than in L4.

<sup>e</sup>Thiol oxidation status of peptides of young adults (day 2).

<sup>f</sup>Thiol oxidation status of peptides in 15-day-old adults.

<sup>g</sup>Tissue localization according to WormBase (<http://www.wormbase.org/>); U, ubiquitous; BW, body wall muscle; M, muscle; N, neuron; I, intestine, P, pharynx; H, hypodermis; G, gonads; R, reproductive system.

<sup>h</sup>Cysteine identified to be peroxide sensitive in *C. elegans* (Kumsta et al., 2010).

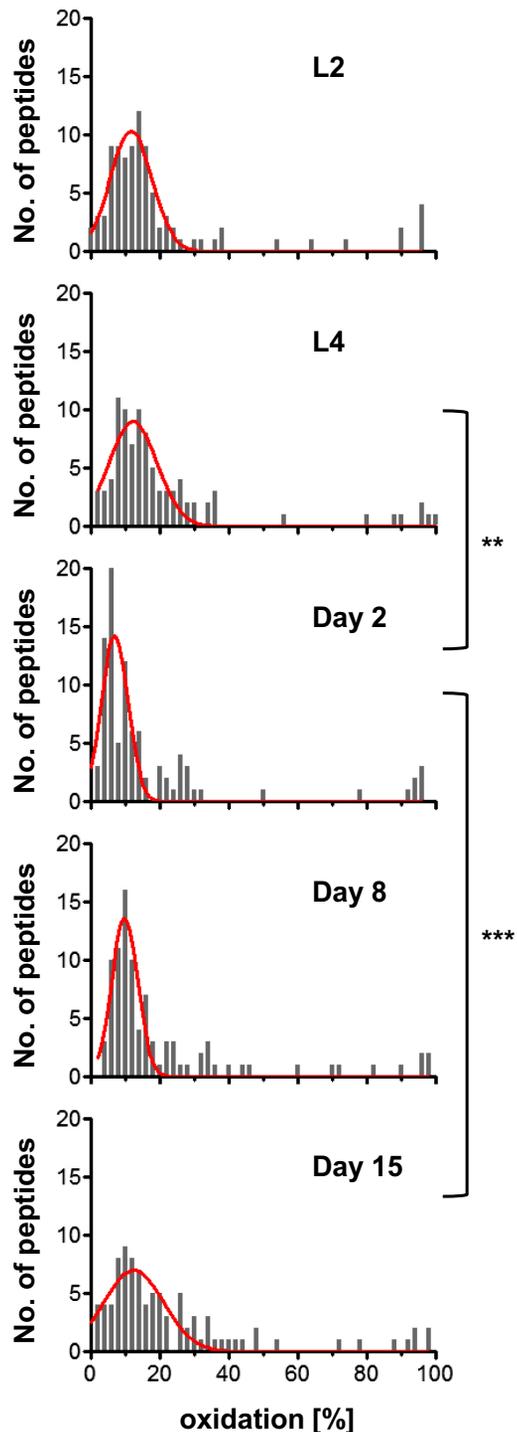
<sup>i</sup>Protein identified to be peroxide sensitive in *C. elegans* (Kumsta et al., 2010).

<sup>j</sup>Protein identified as redox stress sensitive in other systems (Joe et al., 2008; Maeda et al., 2004).

conducted our differential OxICAT thiol trapping. We were able to reproducibly determine the redox status of many of our previously identified peptides at each time point, with the exception of proteins that are either not expressed during early development (e.g., vitellogenin) or whose expression decreases as the worms age (e.g., lysozyme).

We restricted our subsequent analysis to those ~100 protein thiols, which were reproducibly identified at all time points

tested, and plotted them against their respective average oxidation state at each time point (Figure 2). We thus generated histograms that allowed us to directly compare the in vivo oxidation state of a common set of protein thiols during the life span of wild-type *C. elegans* (Table S2). In agreement with the free radical theory of aging, we found that the oxidation status of protein thiols generally increased with age. This was particularly noticeable in aged worms (day 15), which had significantly more



**Figure 2. Monitoring Thiol Oxidation during the Life Span of *C. elegans***

The oxidation status of protein thiols in a synchronized population of wild-type N2 worms cultivated at 15°C was determined at the larval stage L2 and L4 as well as at days 2, 8, and 15 of adulthood. Shown is the frequency distribution of average oxidation levels determined for all of the protein thiols that were reproducibly identified for each time point. The histograms were fitted to a single or double Gaussian model (red line) using GraphPad Prism, with  $R^2$  values of 0.8–0.9. The complete list of proteins and their respective oxidation

oxidized protein thiols ( $p < 0.001$ ) than young adults (day 2). To our surprise, however, we also observed a significantly higher level of thiol oxidation in developing L4 larval worms. This elevated overall protein oxidation significantly decreased ( $p < 0.01$ ) upon entering the reproductive period (day 2). These results suggest that animals are exposed to elevated levels of oxidants early in life, as part of either ROS signaling, increased metabolic activity, or a combination thereof. It is of note that not all protein thiols that we found to be more oxidized during development are also more oxidized in aging worms and vice versa, suggesting more localized oxidation events or antioxidant activities during development and aging.

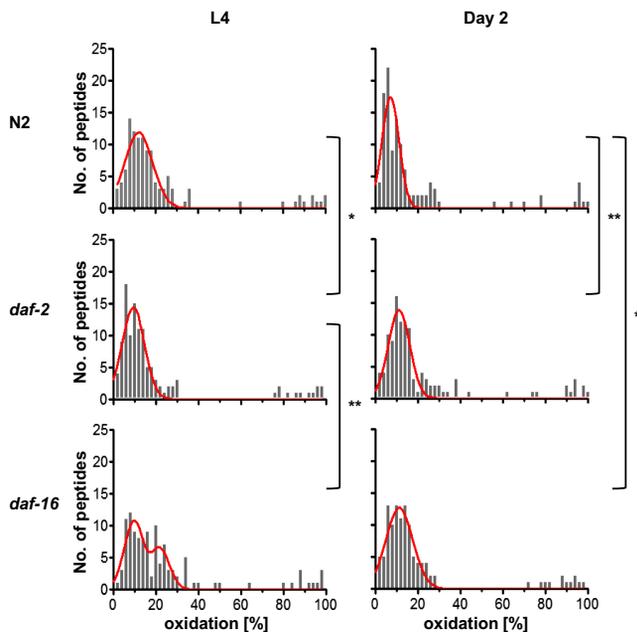
Some of the most significantly oxidized protein thiols during development in *C. elegans* compared to young adult worms were found in the Hsp70 homolog HSP-1, the protease neprylisin, MEL-32, and UNC-87 (Table 1). Many of these proteins were also found highly oxidized in aging *C. elegans*. Proteins that were heavily oxidized in aging organisms versus young adults included the phosphoinositide 3 kinase AGE-1, NDPK, and translation initiation factor 5A. Many of our identified proteins (e.g., HSP-1, UNC-87, ribosomal proteins S28a, S12, L22, translation initiation factor 5A) have previously been observed to contain oxidation sensitive cysteines, either in peroxide-treated *C. elegans* (Kumsta et al., 2010) or in other oxidatively stressed eukaryotic organisms (Fiaschi et al., 2006; Joe et al., 2008) (Table 1). Several proteins, however, have not been shown before to contain oxidation-sensitive cysteines, including AGE-1, which when deleted confers life span extension (Dorman et al., 1995).

Localization analysis (<http://www.wormbase.org/>) of the proteins that showed the most significant oxidation (Table 1) indicated that many of these proteins are ubiquitously distributed. We did observe that a number of our proteins that become substantially oxidized in developing and/or aging worms are exclusively localized to the intestine (neprylisin, vitellogenin-3, vitellogenin-6) or are localized to the intestine and one other cell type (MEL-32, AGE-1) (Table 1 and Table S1). Moreover, UNC-87 and DIM-1, which are both substantially oxidized during development and aging, are localized exclusively to the body wall muscle cells. These results suggest that, at a minimum, increased ROS production occurs in body wall muscle cells and the intestine, cell types known for their high metabolic activity and involvement in aging (Libina et al., 2003).

#### ILS Pathway Mutants Are Affected in Oxidative Stress Recovery

To assess whether the onset, extent, and pattern of endogenous ROS accumulation is altered in mutant strains that show either shortened or prolonged life span, we decided to conduct our OXICAT experiments in worms defective in the ILS pathway. The ILS pathway is evolutionarily conserved, and genetic manipulation of the signaling cascade has been shown to affect the life span of *C. elegans* as early as in young adults (Dillin et al., 2002a).

states can be found in Table S2. Statistical analysis was done using Kruskal-Wallis one-way analysis of variance followed by Dunn's post test. \*\* $p < 0.01$ ; \*\*\* $p < 0.001$ .



**Figure 3. Protein Oxidation during the Life Span of WT, *daf-2*, and *daf-16* Worms**

Relative distribution of protein thiol oxidation in synchronized populations of wild-type N2, *daf-2*, and *daf-16* mutant strains during development (L4) and early adulthood (day 2). Protein oxidation was determined using OxICAT (complete list of proteins and their oxidation states at time points L2, L4 and days 2, 8, and 15 can be found in Table S2). Shown is the frequency distribution of average oxidation levels determined for all of the protein thiols that were reproducibly identified for each time point. The histograms were fitted to a single or double Gaussian model (red line) using GraphPad Prism, with  $R^2$  values of 0.8–0.9. Statistical analysis was done using Kruskal-Wallis one-way analysis of variance followed by Dunn's post test. \* $p < 0.05$ ; \*\* $p < 0.01$ .

DAF-2, the insulin/IGF-1 receptor, negatively regulates the forkhead transcription factor DAF-16, which controls expression of numerous antioxidant genes (Murphy et al., 2003). Reduced DAF-2 function promotes DAF-16-mediated gene expression and increases life span (Kenyon et al., 1993), while deletion of DAF-16 reduces life span (Lin et al., 2001).

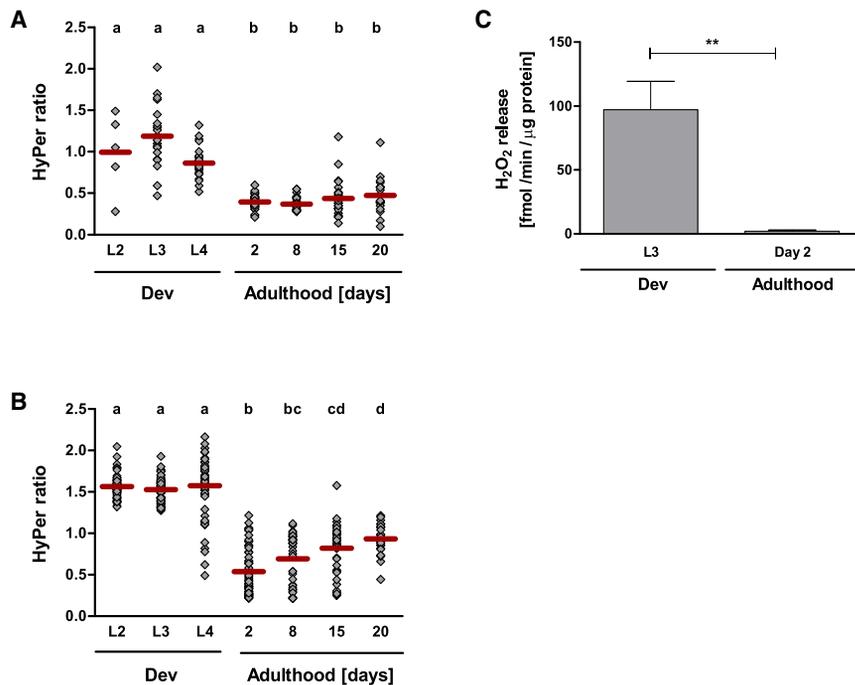
We synchronized worms and took aliquots at the same time points as in the previous experiments. Many of the peptides that we identified in wild-type N2 were also identified in *daf-2* and *daf-16* mutant worms, particularly at early stages in *C. elegans* life, making a statistical analysis of the protein thiol oxidation status possible for worms transitioning from development to early adulthood (Table S2) (Figure 3). We did not observe any significant difference in the overall oxidation status of protein thiols isolated from *daf-2* or *daf-16* worms at the early larval stage L2, suggesting that disruption of the ILS pathway during early development does not globally affect cellular redox conditions (Table S2) (Figure 3). However, differences between the strains were significant by the time *C. elegans* reached the last larval stage (i.e., L4). In the long-lived *daf-2* mutants, protein thiol oxidation was already shifted toward lower levels, whereas in the short-lived *daf-16* deletion strain, protein thiols remained in a more oxidized state. Neither *daf-16* nor *daf-2* mutants changed

their oxidation status significantly upon transition from the L4 state to the young adult state, suggesting that lack of DAF-16 prevents the worm's recovery from high oxidant levels encountered during development. It is of note that no statistically significant difference was observed between the protein thiol oxidation status of wild-type and mutant worms at either day 8 or day 15 (Table S2). This, however, could be due to the fact that at the later time points fewer of our peptides were simultaneously identified in all three strains. Analysis of the oxidation status of the same peptides is, however, crucial for our statistical evaluation.

### Using HyPer to Determine Endogenous Peroxide Levels in *C. elegans*

Many of our proteins, whose thiol redox status was found to vary over the life span of *C. elegans*, have been previously shown to be sensitive to peroxide-mediated thiol oxidation (Table 1). To test whether accumulation of peroxide could be, at least in part, responsible for the observed protein oxidation, we tested the in vivo oxidation status of peroxiredoxin 2 (PRDX-2). This peroxide-detoxifying enzyme undergoes intermolecular disulfide bond formation with a second PRDX-2 molecule as part of its catalytic cycle (Olahova et al., 2008). We found significantly higher amounts of peroxide-induced PRDX-2 disulfides in *C. elegans* larvae as compared to young adults, suggesting that developing animals are indeed exposed to higher peroxide levels (Figure S2). We thus decided to make use of HyPer, a recently developed peroxide-specific sensor protein, which has been previously used to monitor in vivo peroxide levels (Belousov et al., 2006). We reasoned that expressing HyPer in *C. elegans* would allow us to monitor and track peroxide levels in individual worms, and to determine whether the levels of  $H_2O_2$  could possibly correlate with the life span of the animals. The peroxide sensor HyPer consists of a circularly permuted yellow fluorescent protein fused to the  $H_2O_2$ -sensing domain of *E. coli* OxyR (Belousov et al., 2006). The sensor protein possesses two excitation maxima at  $\sim 420$  nm and  $\sim 500$  nm, and a single emission maximum at 516 nm. Upon exposure of HyPer to peroxide, one intramolecular disulfide bond forms within the OxyR domain, causing conformational changes that result in a ratiometric shift. The emission upon excitation at 500 nm increases, whereas the emission after excitation at 420 nm decreases proportionally, leading to an overall increase in the 500 nm/420 nm ratio (i.e., HyPer ratio) with rising peroxide levels.

To use HyPer as an endogenous peroxide sensor in *C. elegans*, we cloned the HyPer gene under the control of the UNC-54 promoter, which targets HyPer expression to the body wall muscle cells, which is one tissue where significant oxidation occurred according to our OxICAT studies (Table 1). We generated a stable transgenic line of wild-type N2 worms expressing the peroxide sensor in the body wall muscle cells (Figure S3A). We backcrossed the strain several times, verified that the expression of HyPer did not affect the life span of the worms (Figure S3B), and confirmed that the HyPer ratio was indeed independent of the amount of HyPer protein expressed (Figure S3C). These features make HyPer well-suited to monitor endogenous peroxide levels over the life span of *C. elegans*.



**Figure 4. Monitoring Endogenous Peroxide Levels during the Life Span of *C. elegans***

The HyPer ratios (A and B) and the H<sub>2</sub>O<sub>2</sub> release (C) were determined as an in vivo readout for endogenous H<sub>2</sub>O<sub>2</sub> levels. To determine the HyPer ratio in the body wall muscle cells of wild-type N2 [*unc-54::HyPer*] (A) or in the head region of wild-type N2 [*rpls1(Prpl-17)::HyPer*] (B) animals at different stages during their life span, worms were cultivated at 15°C and imaged at the indicated time points. Every symbol represents the HyPer ratio of an individual animal; the bar illustrates the average HyPer ratio per day. Experiments were performed a minimum of three times, and representative graphs are shown here (A and B). A one-way ANOVA followed by the Tukey multiple comparison test was performed on the log-transformed HyPer ratio, and means that are not significantly different from each other ( $p > 0.05$ ) share the same letter. The H<sub>2</sub>O<sub>2</sub> release of wild-type N2 animals (C) was measured with the Amplex UltraRed reagent using a H<sub>2</sub>O<sub>2</sub> standard curve. The average H<sub>2</sub>O<sub>2</sub> release of seven independent experiments and the SEM are shown. The means are significantly different ( $p = 0.0011$ , unpaired t test).

### Monitoring Endogenous Peroxide Levels with Temporal and Spatial Resolution

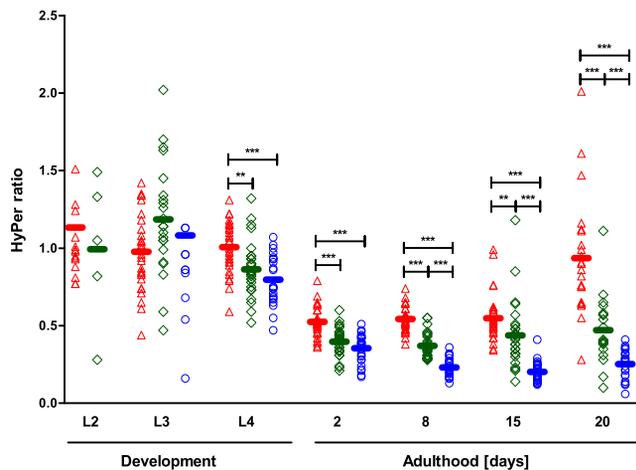
To monitor potential fluctuations in the levels of peroxide during the life span of *C. elegans*, we took about 30 worms of a synchronized population during development (larval stages L2, L3, and L4) and at defined days during adulthood (days 2, 8, 15, and 20) and determined the HyPer ratio for each individual animal. As shown in Figure 4A, we observed high levels of endogenous peroxide during larval development. These results were in good agreement with our OxICAT studies, and suggested that accumulation of peroxide was either directly responsible for increased protein oxidation or indirectly responsible by changing the cellular redox potential toward more oxidizing conditions. Peroxide levels rapidly decreased as the worms reached their reproductive period and remained low during the fertile phase (Figure 4A). To exclude the possibility that the high peroxide levels observed during development were due to the synchronization procedure, which utilizes a hypochlorite solution to release the eggs from gravid adults, we also determined the HyPer ratio in animals that were manually synchronized (Figure S3D). Again, we found a significantly higher HyPer ratio in developing animals as compared to fertile, mature adults, making it unlikely that the high HyPer ratio was caused by the synchronization procedure.

Although our global OxICAT analysis suggested that protein oxidation is likely a system-wide process, we still wanted to test whether other *C. elegans* tissues experience a similar surge in peroxide production during development. We thus conducted an equivalent time course study using a separate transgenic *C. elegans* strain that expresses HyPer under the control of the ubiquitous RPL-21 promoter (Back et al., 2012). This strain was recently used in monoxenic solution studies to assess the peroxide levels in dietary restricted *C. elegans*. We focused on

the endogenous peroxide levels particularly in the head region (i.e., pharynx, neurons) of the worms (Figure S3E), as this region revealed the strongest HyPer fluorescence signal. As before, we found significantly higher peroxide levels during development as compared to early adulthood (Figure 4B). These results strongly suggest that developmental peroxide accumulation indeed involves different tissues of *C. elegans*.

Upon transition to reproductive young adults, peroxide levels appeared to massively decrease and remain low during the majority of the reproductive period. Peroxide levels then slowly increased again as the population aged (Figures 4A and 4B). This increase was not significant in the body wall muscle cells of N2 [*unc-54::HyPer*] worms, potentially because the HyPer fluorescence of immobile worms was very low and could not be quantified. This might have biased the average HyPer ratio of an aged population toward healthier worms. A significant age-accompanying increase in HyPer ratio was, however, observed when we determined the HyPer ratio in the head region of N2 [*rpl-17::HyPer*] worms. These results agreed with recent studies, which showed a global increase in peroxide levels in older worms (Back et al., 2012).

The HyPer sensor allows for a comparative assessment of endogenous peroxide levels but not for their absolute quantification. Moreover, the HyPer ratio has been shown to be also sensitive to pH changes (Belousov et al., 2006). To obtain a more quantitative assessment of endogenous peroxide level in wild-type worms and to exclude the possibility that pH changes are responsible for the observed decrease in HyPer ratio during the L4 to day 2 transition, we decided to monitor the release of peroxide from larval and adult worms using the peroxide-specific Amplex UltraRed reagent (Molecular Probes), which is pH insensitive between pH 5 and pH 10. As peroxide is freely diffusible, the release of peroxide from *C. elegans* should, at



**Figure 5. Hydrogen Peroxide Levels in Wild-Type N2, Short-Lived *daf-16*, and Long-Lived *daf-2* Mutants during Development and Adulthood**

The  $H_2O_2$  sensor HyPer was used to monitor endogenous  $H_2O_2$  levels in wild-type and mutant worms. Every symbol represents the HyPer ratio of an individual animal; *daf-16* [*unc-54::HyPer*] is shown in red, N2 [*unc-54::HyPer*] in green, and *daf-2* [*unc-54::HyPer*] in blue, and the bar depicts the average HyPer ratio per strain and day. The HyPer expression levels in *daf-2* [*unc-54::HyPer*] L2 larvae were too low to allow accurate quantification of the HyPer ratio. Two data points for *daf-2* [*unc-54::HyPer*] L3 larvae are outside the axis limits. Experiments were performed at least three times, and a representative graph is shown. A one-way ANOVA followed by the Tukey multiple comparison test was performed on the log-transformed ratios to compare the means between genotypes within a day. \*\* $p < 0.01$ ; \*\*\* $p < 0.001$ .

a minimum, correlate to the peroxide levels within the worms (Zarse et al., 2012). As before, wild-type animals were synchronized, and the peroxide release of developing animals (L3) and young adults (day 2) was compared over a 1 hr time period. Later time points in life were not assessed by this method, as Amplex UltraRed measurements had to be done in parallel to allow direct comparison of peroxide release. This, however, involved time-delayed cultivation of the strains, which becomes more error prone the longer the time period between start of cultivation and measurement. As shown in Figure 4C, the peroxide release of worms during development was more than 40-fold higher than of young adult worms, indicating that peroxide production and/or secretion of  $H_2O_2$  is dramatically increased in developing animals. These results indicate that peroxide production is strongly enhanced in developing *C. elegans* followed by a rapid cease in production and/or increased clearance as animals reach their reproductive period. These studies, together with our HyPer measurements, strongly suggest that *C. elegans* experiences ROS accumulation at least twice in life, during development and aging.

#### Correlation between Early Oxidative Stress Recovery and Life Span

Our OxICAT analysis revealed a general increase in the thiol oxidation status of protein during late development and also suggested that differences exist in the extent to which animals defective in the ILS pathway might be able to restore redox

homeostasis. To address this aspect in more detail, we generated HyPer-expressing *daf-2* and *daf-16* strains. As before, we imaged aliquots of synchronized populations during development and adulthood to compare the HyPer ratios between the three strains. In agreement with our OxICAT analysis, we did not detect any significant differences in HyPer ratios between wild-type N2 [*unc-54::HyPer*], *daf-16* [*unc-54::HyPer*], and *daf-2* [*unc-54::HyPer*] mutant animals in early development (L2 and L3 larvae) (Figure 5). Animals appeared to be exposed to a significant bolus of  $H_2O_2$  in their body wall muscle cells (Figure 5), implying that at this stage, signaling through the ILS pathway is not involved in ROS generation or ROS detoxification. However, a clear correlation between the time of recovery from oxidative stress and the strain background was detected. By day 2, *daf-2* mutant animals had reached significantly lower steady-state levels of peroxide than the short-lived *daf-16* mutants, and they maintained these low peroxide levels at least until day 20 of their life span. Worms lacking DAF-16, however, never fully recovered from the oxidative stress levels encountered during late development, and peroxide levels remained significantly higher throughout their entire adult life span. Peroxide levels increased dramatically with age, eventually reaching the same levels as those observed during development (Figure 5). These results suggest that the ability to deal with and recover from oxidative stress encountered at very early stages in life might affect redox homeostasis during adulthood and might correlate with the life span of *C. elegans*.

#### DISCUSSION

In this study, we used a set of quantitative tools to determine at what point in life and to what extent multicellular organisms like *C. elegans* are exposed to what type(s) of ROS. We first used the redox proteomics technique OxICAT, which provides a quantitative readout of the thiol oxidation status of proteins. The decision to use OxICAT was based on previous redox proteomic studies that showed that typically 10%–30% of identified thiol-containing proteins get more thiol oxidized in response to increased levels of specific oxidants and/or a more oxidizing redox potential (Held and Gibson, 2012). These proteins contain cysteines that either are exquisitely sensitive to oxidants, such as the active site cysteine of peroxiredoxin, or contain disulfide bonds with redox potentials that are close to the physiological redox potential and thus rapidly adjust their oxidation status to the prevailing redox environment. We hence reasoned that quantitative analysis of the cellular thiol redox proteome will potentially serve two purposes: to function as an endogenous monitoring device of cellular redox conditions and, if changes are observed, reveal the subset of proteins and/or pathways that might be affected by oxidation. Potential shortcomings of this approach include data representation and statistics, as quantitative mass spectrometric techniques are still error prone, and the same peptides are not necessarily identified in all samples at all times. The last aspect is particularly crucial, as distinct protein thiols typically respond differently to changes in their redox environment. This, however, might decrease the number of peptides below the point of statistical analysis. In response to these challenges, we took a histogram approach, simply comparing the redox

status of all those protein thiols that we identified reproducibly at all time points. By using this approach, we found that the redox proteome of wild-type *C. elegans* is statistically significantly more oxidized during development and during aging than during the reproductive phase of young adults. Many of the identified proteins that change their oxidation status during development, adulthood, and aging have been previously shown to be peroxide sensitive (Kumsta et al., 2010). Although this finding suggested that increased peroxide levels might be responsible for the increased oxidation of these proteins, it did not serve as proof, particularly as not all previously identified peroxide-sensitive *C. elegans* proteins were found to be oxidized. To independently test the peroxide levels in developing larvae and young adults, we thus (1) monitored the oxidation levels of peroxide-specific peroxiredoxin, (2) determined the in vivo oxidation ratio of the H<sub>2</sub>O<sub>2</sub>-specific sensor protein HyPer, and (3) quantified the peroxide release using the peroxide-specific Amplex UltraRed reagent. All three methods concurred with our OxICAT results, and showed that animals accumulate high levels of peroxide during development in contrast to very low level of peroxide during early adulthood. Importantly, we found that peroxide levels differed significantly in the ILS pathway mutants *daf-2* and *daf-16*, a difference that first became evident in late development and persisted throughout adulthood. These results support the exciting possibility that animals with improved ability to recover from early oxidative stress might have significant advantages later in life.

Our observations concur with previous studies in *C. elegans* and rodents, which led the authors to conclude that events early in life might dictate life span (Ben-Zvi et al., 2009; Dillin et al., 2002b; Sun et al., 2009). Morimoto and coworkers, for instance, reported recently that the capacity to maintain a functional proteome (i.e., proteostasis) decreases with age and that the onset of the proteostasis collapse, which is particularly obvious in muscle cells and neurons, becomes apparent as early as day 2 or 3 of adulthood (Ben-Zvi et al., 2009). Overexpression of stress transcription factors such as heat shock factor HSF-1 or DAF-16 delayed the collapse and in turn increased life span, whereas deletion of either of these factors accelerated the collapse and decreased life span. It is well known that proteins are one of the main cellular targets of ROS. Once nonspecifically modified by ROS, they often aggregate and require removal by the proteasome (Bader and Grune, 2006). Protein aggregates that are resistant to proteolytic degradation can accumulate in the cell, interfere with basic cellular functions, and ultimately lead to cell death. Based on our results showing that oxidative stress precedes the proteostasis collapse, it is tempting to speculate that the observed lack of recovery from developmental oxidative stress in *daf-16* deletion mutants might contribute to the accelerated collapse of proteostasis and to shorter life span. Conversely, rapidly restoring the redox conditions and keeping ROS levels low, as observed in *daf-2* mutants, might delay the collapse of proteostasis.

Many processes have the potential to increase the levels of peroxide during development. One process that might contribute to increased ROS levels specifically in *C. elegans* is larval molting, as it has been demonstrated that the dual oxidase DUOX, a H<sub>2</sub>O<sub>2</sub>-generating enzyme, is required for correct cuticle

formation (Edens et al., 2001). As peroxide is freely diffusible, this enhanced peroxide production might not only contribute to the massively increased levels of secreted peroxide that we observed during *C. elegans* development (Figure 4C) but might also contribute to elevated peroxide levels in tissues bordering the hypodermis. In addition, increased ROS levels are likely also caused by higher metabolic rates, which have been reported to occur specifically during *C. elegans*' development and are reflected by a peak in ATP generation between the L2 and L4 larval stage (Wadsworth and Riddle, 1989). Moreover, oxidants have been shown to play crucial roles as second messengers in signal transduction, playing a role in development and differentiation (Cross and Templeton, 2006).

Our in vivo redox sensors revealed that *C. elegans* has evolved effective mechanisms to recover from the developmental oxidative burst by the time reproductive age is reached. At least one pathway that we identified to be involved in this recovery process is the ILS pathway (Kenyon, 2010), which functions to mediate stress resistance and life span in adult *C. elegans* (Dillin et al., 2002a). The long-lived *daf-2* mutant lacks the negative regulation of the transcription factor DAF-16, which positively controls expression of stress response genes like superoxide dismutase (SOD) and catalase and thus mediates increased antioxidant capacity. It is interesting to note that we did not observe any significant differences in early developmental ROS levels between short- and long-lived ILS mutants, suggesting that at this stage, DAF-16-mediated expression of antioxidant genes might not occur, thus enabling ROS signaling. It is hence tempting to speculate that growth pathways such as the ILS pathway might support growth in part by allowing increased ROS signaling to occur. We did, however, find a significant difference in the ability of *daf-16* mutants to recover from oxidative stress and to achieve the low ROS levels that appear to accompany adult life span. While *daf-2* mutants recovered from oxidative stress during early adulthood and maintained low ROS levels throughout much of their mature life, *daf-16* mutants failed to fully recover and showed significantly increased ROS levels throughout their life. These results are in good agreement with ILS timing studies, which showed that the ILS pathway influences life span particularly when disrupted in the early days of adulthood (Dillin et al., 2002a). Other antioxidant systems that might play a role in the detoxification of peroxide upon transition from development to adulthood are PRDX-2, which has been previously shown to promote *C. elegans*' recovery from exogenous peroxide treatment (Kumsta et al., 2010), and catalases (CTL-2). Deletion of either one of these two main peroxide-detoxifying enzymes causes progeric phenotypes that become apparent as early as in young adults, and significantly shortens the life span of *C. elegans* (Kumsta et al., 2010; Petriv and Rachubinski, 2004). In contrast, absence of SOD isoforms in *C. elegans* alone or in combination has none or only minor effects on the life span of *C. elegans* (Doonan et al., 2008), and mutants lacking all five SOD isoforms show a normal life span (Van Raamsdonk and Hekimi, 2012). With OxICAT, HyPer, and other related redox sensors, we now have the unique opportunity to precisely track ROS in vivo and to distinguish between ROS that might or might not be relevant for life span.

## Molecular Cell

### Oxidative Stress, an Early Event in Life

A recent study demonstrated that stochastic variances in the expression levels of a stress-inducible gene in early adulthood of *C. elegans* predicts life expectancy in an isogenic population, thus serving as biomarkers of aging (Rea et al., 2005). Our observation that in vivo peroxide levels vary dramatically within an isogenic population of the same chronological age (Figure 4 and Figure 5) supports now the exciting possibility that we have discovered not only another biomarker of aging but possibly one of the main factors contributing to stochastic variances. As transcriptional and epigenetic control mechanisms involve redox-regulated proteins (Cyr and Domann, 2010), early variances in ROS level might play the important role of individualizing gene expression and ultimately determining life span.

#### EXPERIMENTAL PROCEDURES

##### Strains, Culture Conditions, and Life Span Analysis

The *C. elegans* wild-type strain N2 and GR1307 *daf-16(mgDf50)* were provided by the *Caenorhabditis* Genetics Center. Strain CF1041 *daf-2(e1370)* was provided by A. Hsu. The HyPer-expressing strain N2 (*jrls1[Prpl-17::HyPer]*) was provided by B. Braeckman (Back et al., 2012). Strains were cultured on Nematode Growth Media (NGM) agar plates at 15°C using 10<sup>10</sup> cells/ml OP50 as food source and synchronized according to Kumsta et al. (2010). To prevent hatching of progeny, fertile adults were cultured on plates containing 20 mg/l FUDR (Sigma). The L4 molt stage was considered day 0 of adulthood. Animals were considered dead when they did not move or respond to prodding. Animals that crawled off the plate were censored at the time of the event. Generation of the transgenic animals and chromosomal integration are described in detail in the Supplemental Information.

##### Sample Preparation for OxICAT

A synchronized population of wild-type or mutant worms was cultivated at 15°C, and aliquots of worms were taken at the indicated time points. Samples for OxICAT were prepared as previously described (Kumsta et al., 2010; Leichert et al., 2008).

##### Worm Image Acquisition and Image Quantification

Animals of a synchronized population were taken for image acquisition at different time points during larval development and adult life. Worms were mounted on a 2% agarose pad and immobilized using 2 mM levamisole hydrochloride. Up to 60 animals were imaged per day and group. A detailed description of the image acquisition and quantification process can be found in the Supplemental Information.

##### Amplex UltraRed Assay

To monitor the release of H<sub>2</sub>O<sub>2</sub> from *C. elegans* over time, the Amplex UltraRed reagent (Molecular Probes) was used according to the manufacturer's protocol with small modifications. A detailed description of this assay can be found in the Supplemental Information. In brief, synchronized N2 wild-type worms were arrested in L1 and allowed to resume growth in a time-delayed manner, allowing the analysis of peroxide release from developing worms (L3 larva) and young adults (day 2) at the same day. The worms were removed from NGM plates, washed with M9 buffer and then with 1× reaction buffer (Molecular Probes) supplemented with 0.05% Triton X-100, and resuspended in 1× reaction buffer/0.05% Triton X-100. Worm solution (50 μl) was assayed in triplicates according to the manufacturer protocol, and peroxide release per minute was calculated based on a H<sub>2</sub>O<sub>2</sub> standard curve and normalized to the protein concentration.

##### Statistical Analysis

GraphPad Prism 5 (Version 5.01) was used for statistical analysis. Life span data were analyzed using the log-rank test (Mantel Cox) or the Gehan-Breslow-Wilcoxon test; p values < 0.05 are considered significant. The HyPer ratio values were log-transformed and analyzed with one-way ANOVA followed by

the Tukey multiple comparison test. The mean values from the PRDX-2 redox state and the Amplex UltraRed experiments were compared with an unpaired t test with \*p < 0.05 and \*\*p < 0.01. OxICAT data were analyzed using Kruskal-Wallis one-way ANOVA followed by Dunn's post test.

#### SUPPLEMENTAL INFORMATION

Supplemental Information includes three figures, two tables, Supplemental Experimental Procedures, and Supplemental References and can be found with this article at <http://dx.doi.org/10.1016/j.molcel.2012.06.016>.

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